

Total Synthesis of a Glyoxalase I Inhibitor and Its Precursor, (-)-KD16-U1

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Abstract: A glyoxalase I inhibitor and (-)-KD16-U1 have been synthesized from D-ribonic acid γ -lactone through SnCl4-promoted cyclization of a phenylsulfonyl enol silyl ether and regioselective introduction of a hydroxymethyl group. © 1997 Elsevier Science Ltd. All rights reserved.

A glyoxalase I inhibitor (9) was isolated in 1975 from the culture broth of Streptomyces griseosporeus by Umezawa and co-workers.¹⁾ The absolute structure was determined by chemical studies and X-ray analysis.²⁾ Its precursor, (-)-KD16-U1 (8), had been already isolated in 1974 from the culture broth of Streptomyces filipinensis by a chemical screening method developed in our laboratories.³⁾ and converted to the aforementioned glyoxalase I inhibitor by treatment with crotonic acid and BF3-Et2O.⁴⁾ The glyoxalase system. which consists of glyoxalase I, glyoxalase II and reduced glutathione, catalyzes the conversion of α-keto aldehydes to α-hydroxy acids.¹⁾ The glyoxalase I inhibitor (9) has also been reported to exhibit antitumor activities. 1) The structures and bioactivities of these compounds 8 and 9 have attracted our attention because of our program in developing novel methodology for the preparation of densely-functionalized carbocycles from carbohydrates.^{5,6)} The first synthesis was achieved by Vasella's group in which methyl α-D-glucopyranoside was effectively used as a starting material.⁷) Recently, the SnCl4-promoted aldol-like cyclization of phenylsulfonyl enol silyl ethers containing a dimethyl acetal has been explored extensively in our laboratories. 6) This transformation is ideally suited to the synthesis of carbocycle-containing natural products and carbasugars, since the core skeleton arises after appropriate replacement of the phenylsulfonyl group. As an illustration of these strategies, we recently disclosed the total synthesis of (-)-PF1092s.⁵⁾ Herein, we describe the novel synthesis of (-)-glyoxalase I inhibitor (9) and its precursor, (-)-KD16-U1 (8).

In our retrosynthetic analysis, it was proposed that (-)-KD16-U1 (8) could be obtained from the

corresponding \alpha-phenylsulfonyl cyclohexenone 6 through the Michael type addition of tributylstannyl-lithium followed by trapping with formaldehyde and desulfonylation. 8) The cyclohexenone 6 would arise from the enol silyl ether containing the dimethyl acetal 5, which originates from one-step opening of the phenylsulfonylmethyl furanose 4. Thus, the starting material simplifies to commercially available D-ribonic acid γ-lactone.

In practice, the O-trityl derivative 19) was silvlated, followed by reductive de-O-tritylation to give the alcohol 2 (mp 116°C, [α]D +46°)10). Pfitzner-Moffatt oxidation and acetal formation gave 3 (mp 93°C, [α]D +14°). Reaction of 3 with lithiated methyl phenyl sulfone afforded the furanose 4 (mp 136°C, $[\alpha]D + 22^\circ$). This was silvlated to produce, as expected,⁵⁾ the labile enol silvl ether 5 having a simultaneously silvlated hydroxy group (mp 108°C, [a]D +37°). The SnCl4-promoted cyclization of 5 resulted in the formation of the cyclohexenone 6 (mp 197°C, [\alpha]D -120°). Addition of tributylstannyl-lithium⁸) to 6 was followed by trapping of the intermediary β-tributylstannyl sulfone with formaldehyde. This reaction gave an adduct which, upon treatment with silica gel, was converted through β -elimination of the tributylstannyl group to the desired α hydroxymethyl-cyclohexenone 7 (oil, [α]D -72°). De-O-silylation with 90% TFA afforded 8 [mp 114°C, [α]D -166°(H2O)], which was identical with the natural (-)-KD16-U1 (8) in all respects.³)

The synthetic (-)-KD16-U1 (8) was treated with crotonic acid and BF3-Et2O, as previously reported in our laboratories.⁴⁾ to give the selectively acylated product 9 [mp 181°C, [α]D -111°(MeOH)] identical with the natural glyoxalase I inhibitor (9). 1)

Adaptation of this synthetic strategy for the construction of other cyclohexenones is under investigation.

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REFERENCES AND NOTES

- Takeuchi, T.; Chimura, H.; Hamada, M.; Umezawa, H.; Yoshka, O.; Oguchi, N.; Takahashi, Y.; Matsuda, A. J. Antibiot., 28, 737-742 (1975).
- Chimura, H.; Nakamura, H.; Takita, T.; Takeuchi, T.; Umezawa, H.; Kato, K.; Saito, S.; Tomisawa, 2 H.; Iitaka, Y. J. Antibiot., 28, 743-748 (1975).
- Tatsuta, K.; Tsuchiya, T.; Mikami, N.; Umezawa, S.; Umezawa, H.; Naganawa, H. J. Antibiot., 27, 3. 579-586 (1974).
- 4.
- Umezawa, S.; Takita, K.; Takeuchi, T.; Umezawa, H.; Takita, T. Chem. Abstr., 87, 184080b (1977). Tatsuta, K.; Niwata, Y.; Umezawa, K.; Toshima, K.; Nakata, M. Tetrahedron Lett., 31, 1171-1172 5. (1990); Carbohydr. Res., 222, 189-203 (1991).
- Tatsuta, K.; Yasuda, S.; Kurihara, K.; Tanabe, K.; Shinei, R.; Okonogi, T. Tetrahedron Lett., 38, 6. 1439-1442 (1997).
- Mirza, S.; Molleyres, L.-P.; Vasella, A. Helv. Chim. Acta, 68, 988-996 (1985). 7.
- Ochiai, M.; Ukita, T.; Fujita, E. J. Chem. Soc. Chem. Commun., 1983, 619-621 (1983).
- Ireland, R. E.; Anderson, R. C.; Badoud, R.; Fitzsimmons, B. J.; McGarvey, G. J.; Thaisrivongs, S.; Wilcox, C. S. J. Am. Chem. Soc., 105, 1988-2006 (1983).
- 10. Optical rotations were measured in CHCl3 using a 0.5 dm tube at 22°C, unless otherwise noted. Significant ¹H-NMR spectral data (in CDCl₃; 270, 400 and 500 MHz, δ; TMS=0, unless otherwise
 - significant ²H-NMR spectral data (in CDC13, 270, 400 and 300 MHz, 8, 1M3=0, diffess otherwise noted) are the following. 2: 1.90(1H, t, J=5.6Hz), 3.51(1H, ABq, J=3.0, 4.5, 5.6Hz), 3.96(1H, ABq, J=3.0, 4.0, 5.6Hz), 4.45(1H, dd, J=3.0, 4.5Hz), 3: 3.47(3H, s), 3.50 (3H,s), 4.23(1H, d, J=3.0Hz), 4.37(1H, d, J=3.0Hz), 4: 3.46(1H, d, J=15.4Hz), 4.26(1H, d, J=15.4Hz), 7.56(3H, m), 7.98(2H, m), 5: 3.59(1H, d, J=9.0Hz), 3.79(1H, d, J=7.4Hz), 4.32(1H, d, J=9.0Hz), 4.45(1H, d, J=7.4Hz), 5.77(1H, s), 6: 4.18(2H, m), 4.74(1H, dd, J=2.2, 2.0Hz), 7.70(1H, dd, J=2.2, 1.6Hz), 7: 2.28(1H, br), 4.24(2H, m), 6.46(1H, m), 8 (D2O): 4.25(1H, d, J=14Hz), 4.32(1H, d, J=14Hz), 4.47(2H, m), 4.82(1H, m), 6.78(1H, m), 9(DMSO): δ 1.86(3H, dd, J=6.9, 1.8Hz), 4.17(2H, m), 4.55(1H, m), 4.63(1H, dt, J=13.5, 1.5Hz), 4.75(1H, dt, J=13.5, 1.5Hz), 5.07(1H, d, J=3.3Hz), 5.19(1H, d, J=5.5Hz), 5.37(1H, d, J=7.0Hz), 5.92(1H, dq, J=15.6, 1.8Hz), 6.62(1H, m), 6.93(1H, dq, J=15.6, 6.9Hz).